## **SHORT REPORTS**

# VANADIUM IN THE BROWN SEAWEED, DESMARESTIA FIRMA

BRIAN MAXSTEAD COCKERILL, PAUL FINCH and ELIZABETH PERCIVAL

Department of Chemistry, Bourne Laboratory, Royal Holloway College, Egham Hill, Egham, Surrey, TW20 0EX, U.K.

(Received 10 July 1978)

Key Word Index—Desmarestia firma; brown seaweed; alga; vanadium.

Both Desmarestia firma and D. ligulata contain free sulphuric acid (6% of the dry weight in the former species) [1]. No reason for this acid content has so far been discovered. Both species grow below low tide level. Ascidians which grow submerged also contain free sulphuric acid (up to 1.0 molar), and it is considered [2] that this is connected with a high vanadium content (1300 ppm). It has been suggested [3] that the highly acidic environment results in the vanadium oscillating between the oxidation states of 3 and 4 using oxygen gas as the oxidant. To determine if vanadium played a similar role in D. firma, the seawced was analysed for vanadium.

## RESULTS AND DISCUSSION

The concentration of vanadium in the 5 samples varied from 1.50 to 2.50 ppm. Work by Yamamoto et al. [4] on species of Sargassaceae, Ishigeaceae and Spermatochnaceae found that the vanadium contents of these brown seaweeds varied from 0.82 to 10.5 ppm. Although D. firma does contain some vanadium the quantity is comparable with other seaweeds which are devoid of free sulphuric acid. It is therefore concluded that the sulphuric

acid in D. firma has a role different from that present in ascidians which possess an abnormally high content of vanadium, but could perhaps have the same function with another metal.

### **EXPERIMENTAL**

The sample of *Desmarestia firma* was collected from 10 m depth in False Bay, Cape Town, South Africa, by Richard Simon and his colleagues in March, 1975, and was freeze-dried immediately after removal from seawater. Five different aliquots of weed (5 g each) were analysed for vanadium by the method of Jones and Watkinson [5].

## REFERENCES

- Carlberg, G. E., Percival, E. and Rhaman, O. (1978) Phytochemistry 17, 1289.
- 2. Carlisle, D. P. (1968) Proc. Roy. Soc. B171, 31.
- 3. Gillard, R. D. personal communication.
- Yamamoto, T., Fujita, T. and Ishibashi, M. (1970) Rec. Oceanogr. Works Jpn 10, 127.
- 5. Jones, G. B. and Watkinson, J. H. (1959) Analyt. Chem. 31,

Phytochemistry, 1978, Vol. 17, pp. 2129-2130. © Pergamon Press Ltd. Printed in England

0031-9422/78/1201-2129 \$02.00/0

# SESQUITERPENES FROM THE ESSENTIAL OIL OF ASTRANTIA MAJOR

H. BUURMA, R. Bos\*, D. H. E. TATTJE and J. H. ZWAVING Laboratory of Pharmacognosy, State University, Groningen, Antonius Deusinglaan 2, The Netherlands

(Received 20 May 1978)

Key Word Index—Astrantia major; Umbelliferae; essential oil; sesquiterpenes;  $\beta$ -sinensal;  $\beta$ -sinensol;  $\beta$ -sinensyl acetate.

Astrantia major L., (family Umbelliferae, subfamily Saniculoideae) is a robust perennial found in Central Europe. The plant has previously been investigated for various constituents [1, 2]. However, although the essential oils of many species of the Umbelliferae have been extensively studied, very little is known about those

of A. major or other members of the subfamily Saniculoideae [2]. Since the chemical characters of these oils clearly have a high selective value for chemotaxonomic purposes [3] an investigation into the constituents of the essential oil of Astrantia major was started. This publication deals with the isolation and structural elucidation of three sesquiterpenes, isolated from the essential fruit oil.

<sup>\*</sup> Where reprints are available.

2130 Short Reports

R = CHO  $\beta$ -sinensal  $R = CH_2OH$   $\beta$ -sinensol  $R = CH_2OOC_2H_5$   $\beta$ -sinensyl acetate

The essential oil from the fruit of Astrantia major was divided into several fractions by column chromatography on Si gel. One fraction contained a mixture of sesquiterpenes which was investigated by GC-MS. In this way three liquid sesquiterpenes were isolated and identified as  $\beta$ -sinensal,  $\beta$ -sinensol and  $\beta$ -sinensyl acetate. Their structures were elucidated by spectroscopic methods.  $\beta$ -Sinensal has been identified as a flavour constituent in the peel of the sweet orange, Citrus sinensis L. [4] and has already been synthesized [5-7].  $\beta$ -Sinensol which has been also synthesized [5, 6] and  $\beta$ -sinensyl acetate are now found for the first time as natural compounds. The absolute configurations of these three sesquiterpenes were established with  $^{13}$ C NMR and they were shown to be the trans-trans isomers.

### **EXPERIMENTAL**

Distillation of the essential oil. Fruits of Astrantia major L. (287 g) were submitted to steam distillation with the apparatus for the determination of volatile oils according to the British Pharmacopoeia [8]. Yield 3.40%.

Isolation of sesquiterpenes from the essential oil. The essential oil (9.76 g) was divided into several fractions by Si gel CC (3  $\times$  30 cm) using petrol (bp < 40) containing an increasing percentage of Et<sub>2</sub>O (0  $\rightarrow$  100 %). The fraction with 10 % Et<sub>2</sub>O was investigated by GLC using a 2 m  $\times$  2.3 mm column of Carbowax 20 M on Chromosorb G HP 80-100 mesh; temp. 80-200° (4°/min); flow 30 ml N<sub>2</sub>/min; TC.

GC-MS was performed on a Finnigan 3300 quadrupole computerized system, direct inlet. Electron energy 70 eV; scan speed 2 sec: scan ion source 200.

Spectral data.  $\beta$ -sinensal: The spectra were identical to the literature, IR [5, 6], NMR [4, 5, 6] MS [9] 218 (M<sup>+</sup>), 93 (= 100).

β-sinensol: IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>-1</sup>: 3380, 3120, 3050, 2950, 2890, 1605, 1460, 1395, 1245, 1170, 1080, 1025, 1000, 902, 860. NMR, 60 MHz, CCl<sub>4</sub>: δ 1.68(3H, s, CH<sub>3</sub>—), 1.71 (3H, s. CH<sub>3</sub>—), 2.11, 2.22, 2.29 (4 —CH<sub>2</sub>—), 3.93 (2H. s, —CH<sub>2</sub>O—), 5.01 (2H. br s) 5.08–5.38 (4H), 6.12–6.60 (1H, q). MS  $m/e(\frac{\nu}{2})$  392(1), 40(5), 41(60), 42(4), 43(100), 53(18), 55(33), 57(12), 65(8), 67(33), 68(13), 69(22), 77(16), 79(30), 80(5), 81(18), 84(5), 91(19), 92(8), 93(78), 94(12), 95(11), 105(8), 106(4), 107(12), 109(5), 119(15), 120(18), 121(13), 133(27), 134(5), 147(3), 161(4), 173(1), 189(2), 220(M<sup>-</sup> · .2). 1<sup>3</sup>C NMR (C<sub>6</sub>D<sub>6</sub>): C-1(CH<sub>2</sub>OH) 67.6; C-2 134.1; C-3 124.5 or 123.6; C-4 26.5 or 26.2; C-5 38.9; C-6 134.1; C-7 123.6 or 124.5; C-8 26.2 or 26.5; C-9 30.9; C-10 145; C-11 138.3; C-12 114.9; C-2 Me 13.0; C-6 Me 15.4; C-10 = CH<sub>2</sub> 112.1 ppm. The trans-trans configuration was concluded from the following data: cis-nerolidol C<sub>5</sub> 31.9; C<sub>6</sub>-Me 22.5 ppm: transnerolidol C<sub>5</sub> 39.7; C<sub>6</sub>-Me 16.0 ppm. In our compound C<sub>5</sub> 38.9; C<sub>6</sub> 15.4 ppm.

β-sinensyl acetate: IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>-1</sup>: 3090, 2975, 2930, 2860, 1745, 1596, 1460, 1440, 1390, 1378, 1358, 1230, 1045, 1020, 986, 950, 892, 842. NMR 60 MHz, CCl<sub>4</sub> δ: 1.56 (3H, s) 1.61 (3H, s), 2.04 (3H, s), 2.11, 2.22, 2.29 (4-CH<sub>2</sub>—), 4.41 (2H, s, —CH<sub>2</sub>O—), 4.99 (2H, br s), 5.02–5.48 (4H, 2-CH=...=CH<sub>2</sub>), 6.12–6.60 (1H, q, —CH=CH<sub>2</sub>). MS  $m/e("_0)$ : 41(27), 43(100), 53(9), 55(16), 65(5), 67(22), 68(10), 69(5), 77(8), 79(18), 81(10), 85(5). 105(6), 107(8), 119(9), 120(8), 133(14), 145(2), 146(3), 159(2), 173(2), 187(3), 202(1), 262(M<sup>-7</sup>, 2).

Acknowledgements.—The authors wish to thank Mr. J. F. Visser for the cultivation of A. major in our experimental garden at Buitenpost, Dr. A. P. Bruins for performing GC-MS and Drs. G. Sipma and Mrs. J. Kloosterman from Polak's Frutal Works, Amersfoort (the Netherlands) for performing <sup>13</sup>C NMR.

#### REFERENCES

- 1. Harborne, J. B. and Williams, C. A. (1972) Phytochemistry 11, 1741.
- Hegnauer, R. (1973) Chemotaxonomie der Pflanzen Vol. 6, pp. 563, 599-601. Birkhaüser, Basel, Stuttgart.
- 3. Harborne, J. B. and Heywood, V. H. (1969) Phytochemistry 8, 1963.
- Stevens, K. L., Lundin, R. E. and Teranishi, R. (1965)
  J. Org. Chem. 30, 1690.
- 5. Büchi, G. and Wüest, H, (1967) Helv. Chim. Act. 50, 2440.
- 6. Bertele, G. and Schudel, P. (1967) Helv. Chim. Act. 50, 2445.
- 7. Thomas, A. F. (1967) Chem. Commun. 947.
- 8. British Pharmacopeia 1973, A87.
- 9. Thomas, A. F., Willhalm B. and Müller, R. (1969) Org. Mass Spectrom. 2, 223.